

Article

## Advanced Development of Bio-fertilizer Formulations Using Microorganisms as Inoculant for Sustainable Agriculture and Environment – A Review

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any medium, provided the original work is properly cited.

*Abstract*— Conventional types of fertilizer such as chemical and synthetic fertilizers have demonstrated various adverse effects on the environment, crops, and humans. The utilization of plant probiotics as a bio-fertilizer in agriculture has been recognized to benefit the growth of the plant and inhibit the activity of plant pathogens. Traditional formulations of bio-fertilizer have provided insight into the beneficial use of microorganisms in crops. Despite its advantage to the environment, the effectiveness of traditional bio-fertilizer is common as compared to chemical fertilizer. Thus, a variety of bio-fertilizer formulations have been developed to improve the success rate of bio-fertilizer in increasing plant productivity. This review was focused on the development of bio-fertilizer formulation and the potential of bio-fertilizer to substitute chemical fertilizer application. In addition, this research review was also undertaken with a great demand on producing low cost and highly effective fertilizer without harming the environment and humans. Thus, the advantages and disadvantages of each formulation type have also been reviewed, emphasizing the perspective of bio-fertilizer and their suitability as bio-fertilizer as a substitute for chemical fertilizers in sustainable agriculture.

*Keywords*— Bio-fertilizer formulation, plant probiotic, sustainable agriculture

#### I. INTRODUCTION

The crop is one of the main food sources for humans in order to survive. Therefore, the abundant yield of crops such as rice is crucial in maintaining food supply and demand all over the world. However, some crops are likely to suffer from disease or cannot consistently grow due to several factors such as unsuitable environment, lack of nutrients, pest, infection, and climate change [1, 2]. Rice, which is known as a staple food, is consumed all over the world, facing critical threat diseases such as blast, blight, brown spot, narrow brown spot, bacterial leaf streak, and rice ragged stunt virus disease [3,4]. In tomato crops, the most common disease detected using machine learning is the yellow leaf curl virus, bacterial spot, late blight, Septoria leaf spot, spotted spider mite, early blight, and mosaic virus [5]. Meanwhile, the disease that causes gall formation in root tissue in *Brassicae* crops, including *Arabidopsis thaliana* and Chinese cabbage, is known as clubroot disease, which is caused by *Plasmodiophora brassicae* [6].

Most plants or crops require extensive care to prevent them from developing the disease symptoms caused by pathogen infection. As a part of infection strategy, many pathogens modify plant development which causes a broad spectrum of impacts from subtle manipulation of host metabolism to extensive tumor formation. For instance, Blumeria graminis lives in the host epidermis and shows only the subtlest effects on the plant, such as barley, while Ustilago maydis cause a strong effect on its host by developing tumors on above-ground tissues of crops such as corn [7, 8]. Crops rely on macronutrients such as nitrogen, potassium, phosphorus to boost productivity. Therefore, chemical fertilizers have been widely used to overcome these problems. The chemical fertilizer is commonly used in the agricultural industry as it contains more nutrients and can channel nutrients instantly to the crop compared to the organic fertilizer [9, 10]. However, it has been reported to cause adverse effects [11].

The pressure to produce more crop yield has led to the uncontrolled application of fertilizer and pesticides in agriculture. Farmers tend to over-apply chemical fertilizer such as nitrogen, which can cause softening of plant tissue and therefore make the plant more susceptible to disease and infection [12]. In addition, the active ingredients contained in pesticides and chemical fertilizers often leached into the water system, causing water pollution, which could endanger the ecosystems. This issue was supported by a previous study that stated cultivation of rice had been one of the main causes of polluting the environment [13, 14]. Other than that, spraying various types of pesticides without the proper protection may also harm the farmer as they are exposed to toxic chemicals. Later, this may cause skin allergy, breathing difficulties, and other health problems [15]. Microorganism populations in the soil make an important contribution to supply required nutrients to plants and increase resistance toward infection and pathogens [16].

The application of bio-fertilizer could be the alternative way to increase crop productivity in the agricultural sector. Thus, this review was focused on the development of bio-fertilizer formulation and the potential of bio-fertilizer to substitute chemical fertilizer application. This research review was undertaken with a great demand on producing low cost and highly effective fertilizer without harming the environment and humans.

# II. BIO-FERTILIZER AND THEIR RELATIONSHIP WITH HOST PLANT

Colonization of plant-associated microorganisms has been recognized to meet the significant functions in crop development and productivity. Microorganisms (bacteria and fungi) are capable of performing various mechanisms, which are directly essential to increase plant growth and restrain the activity of plant pathogens. Therefore, the plant-microbiome interaction has created a great potential in agriculture where it is conceivable to establish a bio-fertilizer for plants [17]. Biofertilizer refers to the application of beneficial microorganisms in the fertilizer and is also known as microbial-based fertilizer.

Bio-fertilizers have great potential in maximizing and augmenting crop production. Bio-fertilizer has recently gained popularity due to its promising ability to increase yield and promote the growth of plants, including oil palm, paddy, and wheat [18, 19, 20, 21, 22]. Similarly, the inoculation of biofertilizer into crops helps to amend soil fertility and be ecofriendly whilst, at the same time, being cost-effective since the quantity of chemical fertilizer used is minimized [19, 23, 24, 25]. The incorporation of microorganisms as bio-fertilizer has a special impact on the host plant. It is different from any other fertilizer, such as organic fertilizer, as it does not provide nutrients that can be directly taken by the plant. Instead, the bacteria contained in the bio-fertilizer will colonize in the soil and complete certain mechanisms to convert the organic form of nutrient into soluble form for plant uptake.

The concept of beneficial interaction between bio-fertilizer and host plant is important as they promote effective and ecofriendly approaches. Bio-fertilizer can provide essential nutrients for plants and crops such as nitrogen, phosphate, and potassium. For example, nitrogen naturally exists in the atmosphere as nitrogen gas. However, plants are unable to use the natural form of nitrogen directly. Thus, a bio-fertilizer containing nitrogen-fixing bacteria (NFB) will convert it into ammonia form (NH<sub>3</sub>) for plant uptake. This process also applies to other major nutrients required by the plant. The idea of developing bio-fertilizer was meant to improve crop yield profitability and simultaneously diminish the reliance on chemical fertilizer. It is well known that chemical fertilizer poses a dangerous threat to humans and the environment over the long haul. Thus, commercializing the use of bio-fertilizer would be a better choice to protect the environment, ecosystem, and humans.

Plant growth-promoting rhizobacteria (PGPR), which contain bio-fertilizer, are important for plant health. PGPR acts towards plants by using different mechanisms of action. The simple mechanism of PGPR action is by releasing nutrients such as vitamins, iron, hormones, solubilization of phosphate and ammonia into the soil and then taken up by plants to sustain rapid plant growth [26, 27, 28]. In addition, the interaction between PGPR and plants is supported by cell-to-cell communication, for instance, through quorum sensing, as signaling mechanisms to monitor the surrounding and microbial activities around the rhizosphere [29, 30, 31]. Finally, chemotaxis and chemokinesis reactions through chemotactic agents such as amino acids and sugar are important to increase the mobility of bacteria to establish the interaction between PGPR and plants [26, 32].

#### III. THE USE OF MICROBIOME FOR THE DEVELOPMENT OF BIO-FERTILIZER

The role of microorganisms and their interaction with plants are exploited to produce bio-fertilizer. Most microorganisms used in bio-fertilizer are commonly extricated from rhizosphere soil which is in contact with plant roots [27, 32]. A number of studies have been performed to distinguish between beneficial and harmful rhizosphere bacteria according to their impact on soil fertility, plant growth, and productivity [33, 34]. For instance, members of the genus *Bacillus*, which is classified as plant growth-promoting rhizosphere (PGPR) isolated from tomato rhizosphere, could increase the growth of tomato plants [33]. In addition, cover crops such as pea, rapeseed, and wheat could increase the growth of plantbeneficial bacteria, which could restore declining soil nutrients and control bacterial wilt disease in tobacco fields [34]. Beneficial microorganisms, also known as plant probiotic bacteria, are capable of exhibiting beneficial effects in agriculture such as fixing nitrogen, improving the cycling of nutrients, detoxifying pollutants, and generating bioactive compounds such as hormones and enzymes [35, 36, 37]. Furthermore, the application of various beneficial microbiomes can generate 'induce systemic resistance,' which is able to fight against multiple plant pathogens [36].

The development of bio-fertilizer requires extensive research and proper experimental design to establish highquality bio-fertilizer. As reported in [38], there are five fundamental procedures involved in developing carrier-based bio-fertilizer (Figure 1). The first step is the isolation and identification of plant-associated microorganisms. The second is the preparation of starter culture and inoculum by cultivating the microorganisms, followed by preparation of carrier materials and incorporating microorganisms into carrier materials. The final step is the packaging and storing carrierbased bio-fertilizer [38].

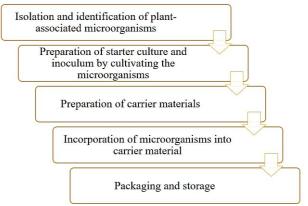


Figure 1. Procedures in developing carrier-based bio-fertilizer

#### A. Selection of Plant Probiotic Strains

Selection of the right strain of bacteria is crucial as it is the primary step in producing bio-fertilizer. A suitable strain of plant probiotic bacteria must be carefully selected to ensure their effectiveness to the host plant. Several criteria need to be considered when selecting the microorganisms. As stated in [24], some of the essential requirements in selecting plant probiotic strains to be used in bio-fertilizer are effectiveness (provide nutrients for plants), competitiveness, being able to initiate plant defense and showing resistance traits against pathogens. In addition, plant probiotic bacteria can be classified based on their functions and nature.

1) Plant Growth-Promoting Bacteria: This type of plant probiotic comprises free-living bacteria that colonize in soil and certain parts of plant tissue to develop a special symbiotic interaction with a host plant. In the interaction phase, the bacteria may carry out direct or indirect mechanisms to boost plant productivity. They can either directly promote the growth by providing the available nutrients or regulating the levels of plant hormones, or indirectly by combating pathogenic agents that have an inhibitory effect on plant development. Plant growth-promoting bacteria (PGPB) have the capacity to synthesize various kinds of antibiotics to inhibit plant pathogens [39]. Among the bacteria that pose functional properties of PGPB are *Rhizobium*, *Azospirillum*, *Anabaena*, *Acetobacter*, *Bacillus megaterium*, *Azolla*, *Pseudomonas*, and *Bacillus polymyxa* [40]. *Pseudomonas putida* strain, for instance, has effectively improved the wheat yield by inhibiting negative impact and reducing stress on the host plant [41].

2) Nitrogen-Fixing Bacteria: Nitrogen fixers are the bacteria that fix atmospheric nitrogen and have interchangeable beneficial interactions with the host plant. The natural form of nitrogen must be converted into ammonia for plant absorption through a process of nitrogen fixation. This process is frequently carried out by nitrogen-fixing bacteria (NFB) that are present in the soil. In this process, nitrogen from the atmosphere enters the soil, and nitrogen fixers fix nitrogen into ammonium ions by releasing nitrogenase enzymes. Several strains of nitrogen-fixing bacteria include Azotobacter sp., Rhizobium sp., Cyanobacteria, and others [42, 43]. Besides that, Azotobacter also can fix a considerable amount of nitrogen in the atmosphere and be able to generate antifungal compounds against plant pathogens. Inoculation of nitrogenfixing bacteria as bio-fertilizer is important to supply adequate nutrients to plants and maintain the fertility of the soil. For instance, the Azotobacter tropicalis strain contributed to a positive impact on maize crops by stimulating a four-fold increase in crop growth [44]. Thus, plants can minimize their dependence on chemical fertilizer due to nitrogen fixer's ability.

3) Phosphate-Solubilizing Bacteria: Phosphorus solubilizers are microorganisms that are able to solubilize insoluble phosphorus to supply soluble phosphorus that can promote plant growth. Phosphate-solubilizing bacteria can perform specific mechanisms to solubilize phosphorus by producing organic acid such as citric acid. This acid induces low pH conditions and therefore results in the dissolution of bound phosphate [42]. The conversion of insoluble phosphorus was mainly performed by carboxyl and hydroxyl groups present in organic acid that chelate the cations bound to phosphate. Phosphorus has an important effect on the photosynthesis, maturity, and disease resistance of the plant. As reported in [45], inoculation of Bacillus megaterium can increase the uptake of phosphorus and tuber size in potato crops. The identified strains of microorganisms that exhibit the ability of solubilizing phosphate are Bacillus subtilis, Pseudomonas sp., Agrobacterium sp., Acetobacter sp., and Azotobacter sp. [40, 46]. Therefore, inoculation of these bacteria will effectively maintain the supply of soluble phosphorus for the plant.

4) Arbuscular mycorrhizae: Mycorrhizae is basically a fungus that establishes symbiotic relationships with the root of the host plant. They attach to the plant root system to absorb nutrients and form a mild type of parasitism that is symbiotic, in which both organisms (plant and fungus) obtain benefit from the association. This can be explained as most plants depend on mycorrhizal fungi to channel moisture and essential nutrients required by the plant, while in return, the plant helps supply the fungus with substances and nutrients produced by the plant. They provide various benefits to the host plant, such as aiding in the major nutrient uptake and reducing the vulnerability to water stress [47]. Certain mycorrhizal fungi are capable of forming a protective cover around the root, making the plant more tolerant to drought, extreme temperature, extreme soil acidity, and infection by pathogenic fungi. As reported in [48], they discovered *Rhizophagus fasciculatus* as the best strain to enhance salinity stress of *Casuarina equisetifolia L*. and increase the nutrient content in the plant.

#### B. Selection of Carrier Material

Bio-fertilizer can be classified into several types, such as peat formulation, liquid formulation, and granules formulation [40]. Bio-fertilizers are typically supplemented with the carrier material. However, it can also be formulated without the inclusion of carrier material such as liquid bio-fertilizer. Carrier materials are commonly added into the formulation to improve the potency of bio-fertilizers. The primary role of carrier substances is to provide the inoculated microorganisms with a suitable and appropriate environment to increase biofertilizers shelf life and effectiveness. The inclusion of carrier materials can facilitate the handling and application of biofertilizer as well as enhance the storage condition. In order to develop good quality bio-fertilizer, a proper selection of carrier material is required. In selecting the most suitable carrier material, the following properties need to be considered such as prices, organic matter quality, contaminants, water retention capacity, processing method, friability, and vulnerability. Ideal carrier material should be readily accessible, lower in prices, free from hazardous contaminants, have a water retention capacity of more than 50%, simple to process, be non-clumping material, and be easy to sterilize [49]. In nature, carrier materials may be organic or inorganic. Followings are the types and examples of carrier [50]:-

1) Soils: Inorganic soil, peat, clays, coal, lignite.

2) Plant waste materials: Composts, cellulose, farmyard manure, charcoal, peanut oil, wheat bran, corn cobs, press mud.

*3) Inert materials:* Vermiculite, talc, rock phosphate, perlite, calcium sulfate, alginate beads, polyacrylamide gels.

4) *Plain lyophilized microbial cultures and old dried bacteria:* Can be readily used or may be integrated with a solid carrier.

The selection of carrier substance depends on the desired function and aims to produce the bio-fertilizer formulation, either in granule, pellet, immobilized, or other forms. During the preparation of the carrier, the sterilization process is one of the critical parts to ensure the bio-fertilizer produced is safe from any pathogenic organisms. The type of sterilization methods may be variant depending on the type of carriers used. Gamma-irradiation and autoclaving are the common methods used for carrier sterilization.

### IV. HISTORY ON THE APPLICATION OF BIO-FERTILIZER

Traditional agriculture was practiced a thousand years ago. Traditional farming is based on the thousand years of experience and knowledge practiced by local farmers. The traditional practices have provided insight and basic knowledge in developing scientific expertise and advanced application in agriculture. Traditional formulations made from organic materials or wastes are used as bio-fertilizer to supply essential nutrients to plants [51]. The application of traditional formulations helps to conserve the environment, soil quality as well as microbial diversity in soil. It is important for soil health as it facilitates the colonization of microorganisms and enhances microbial diversity. In most Asian countries, the local farmers tend to create their own traditional bio-fertilizer formulations by incorporating various organic compounds, which then undergo fermentation or composting to boost their activities and functions (Table I).

 TABLE I

 EXAMPLES OF TRADITIONAL BIO-FERTILIZER FORMULATION

Countries	The traditional formulation of bio-fertilizer		
Indonesia	Bio-fertilizers containing inoculants of		
	symbiotically nitrogen fixer bacteria		
	(Bradyrhizobium japonicum) were used to		
	substitute chemical nitrogen fertilizers for		
	soybean cultivation [52].		
South Korea	Wild weed wineberry extract with		
	indigenous microorganisms was used by		
	combining the extract of wild weeds,		
	molasses, salt, and extract of decomposed		
	leaf soil [53].		
Thailand	The extract of the fermented plant was used		
	to stimulate plant growth and serve as a		
	bio-control agent [54].		
India	Panchagavya, the traditional bio-fertilizer		
	formulation, is prepared by fermenting		
	cow-based products such as milk, urine,		
	dung, curd, and clarified butter [55].		
Japan	Traditional formulation known as Bokashi		
	in the 1980s contained effective		
	microorganisms (EM). It has been used as a		
	starter to ferment raw materials such as		
	residue from plant or animal materials and		
	then produced into bio-fertilizer [56].		

A group of researchers [57] has reported a study on a traditional organic formulation that uses organic fertilizer extract with 2% yogurt. The finding showed that the type and concentration of components used directly contribute to the variation of the microbial population on the host plant. The application of traditional formulation has shown a good effect on plant growth. Chemicals are not utilized in this type of traditional farming. Thus, the quality and pH balance of the soil can be maintained. In contrast, it yields an indifferent impact on crop productivity. This may be due to limited information available on the microbial and chemical components contained in it. Following the two world wars, traditional agriculture has become ordinary and indifferent, as, during that period, the expertise in chemistry has significantly improved. Modern formulations of fertilizer such as synthetic fertilizers, chemical fertilizers, and pesticides have been commercialized due to their promising effects on crop production and yield. However, the application of modern fertilizer may cause long-term adverse effects on the environment and diminish the supply of natural resources and soil microbes in agriculture. Therefore, in the past decades, many researchers have formulated various types of bio-fertilizers with extensive and advanced effects on crop growth to preserve the environment and ecosystem.

#### V. ADVANCED DEVELOPMENT OF BIO-FERTILIZER FORMULATION USING BENEFICIAL MICROORGANISM

#### A. Liquid Formulation Bio-fertilizer

A liquid bio-fertilizer is a bio-fertilizer that comprises viable microorganisms in liquid form. This liquid inoculant grows selected microorganism strains without the inclusion of solid carrier material. After screening and identification of effective microbes to be used as bio-fertilizer, the cells will be cultured in a broth medium rich in nutrients [58]. Once the cells have been cultured and propagated on a larger scale, the liquid inoculants are ready to be applied to the seed or host [59].

There are several formulations of liquid bio-fertilizer that have been established by previous researchers (Table II). Liquid bio-fertilizer seems to be an effective bio-fertilizer formulation for many crops, especially for the cultivation of crops without using soil (hydroponic). The development of low-cost bio-fertilizer yet effective is favorable in the agricultural industry. As stated [60], a simple liquid inoculant in sterile water was able to improve the quality and antioxidant level in strawberry fruits. Researchers of the Malaysian Nuclear Agency, as in [61], studied the ability of four broth mediums (sterile distilled water, nutrient broth, Luria-Bertani broth, and Tryptic soy broth) in retaining the viability of plant probiotic cells to produce high shelf life of liquid bio-fertilizer with low production cost. Based on the result, the nutrient broth is the best medium to retain the survivability of bacteria (Phosphate solubilizing bacteria and plant growth-promoting bacteria (PGPB)) compared to other mediums. Similarly, a liquid formulation of several strains of bio-fertilizer with added nutrients such as rock phosphate, K-feldspar, and natural mineral fertilizers has shown extensive growth on vegetables [44].

Apart from that, [62] proposed that treatment of bacterial cells with polymeric additives helps to maintain the viability of bio-fertilizer over long periods of storage. The additives help to enhance the shelf life and adherence of bio-fertilizer to seed [63]. Furthermore, [62] found that the development of liquid bio-fertilizer using a combination of glycerol and polyvinyl pyrrolidone has the longest shelf life and excellent efficiency over 180 days of storage.

#### B. Granules Formulation of Bio-fertilizer

Granules formulation is a type of carrier-based biofertilizer formulated in the form of small like-grain particles. Granules form was first invented in the 1990s [64]. The main principle of this formulation is to create dust-free bio-fertilizer without having concern about powder segregation [65]. Besides that, this formulation plays an important role in ensuring the uniformity of the bio-fertilizer content in each granule particle.

TABLE III LIQUID FORMULATIONS OF BIO-FERTILIZER

Bio-fertilizer/ Strains	Formulation	Result	Ref.
Phosphate solubilizing bacteria (PSB), Plant growth- promoting bacteria (PGPB)	Broth media: Sterile distilled water, Nutrient Broth (NB), Luria-Bertani Broth (LB), Tryptic Soy Broth (TSB)	NB formulations are effective for microbial survival.	[61]
Azotobacter tropicalis, Burkhoderia unamae, Bacillus subtilis	Addition of rock phosphate and K- feldspar	Vegetables' growth improved by seven times.	[44]
Bacillus amylolequefaciens (BChi1), Paraburkholderia fungorum (BRRh- 4)	Sterile water (nutrient- free bacterial suspension)	Enhanced strawberry quality.	[60]
Rhizobium, Azotobacter, Azospirillum, Bacillus megaterium	Cell protectants: Glycerol, Polyvinyl pyrrolidone (PVP), Polyethylene glycol (PEG), Gum Arabic (GA), Sodium alginate (SA)	Glycerol and PVP are the best cell protectants.	[62]

Granule formulation is composed of carrier materials that are moistened with an adhesive and blended with powdered form inoculum [66]. Followings are the basic process in producing granules formulation. After preparing powdered inoculum from the efficient strains of the microbiome, the ideal carrier material is processed into fine powder. The carrier material will be mixed with bacteria inoculum using a mechanical mixer or manually until both components blend homogeneously. In order to produce a grainy-like structure of bio-fertilizer, the homogenous mixture is loaded into a rotary drum granulator, and the particles will be air-dried upon storage. Granules formulation of bio-fertilizer is developed due to its reasonable function and association with the host plant. This formulation can be modified in various ways based on the function and characteristics that one desires to convey (Table III). According to [67], a granular formulation made from legume inoculation is better in terms of nitrogen accumulation, N2 fixation, and nodule amount and weight compared to liquid inoculant. Besides that, [68] acknowledged that granular formulation of phosphate solubilizing Burkholderia with clay, rock phosphate, and rice bran as a carrier had demonstrated high viability of bacteria during storage. Figure 2 shows an example of urea granule coated with castor oil and starch gel and Aspergillus niger as biological activation microorganisms [69].

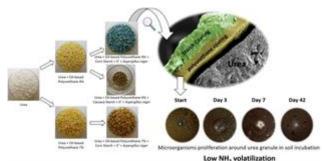


Figure 2. Urea granule bio-fertilizer coated with starch, urethane, and biologically activated by *Aspergillus niger* [69].

TABLE IIIII GRANULE FORMULATIONS OF BIO-FERTILIZER

Bio-fertilizer/	Formulation	Result	References
Strains			
Legume inoculation	Employed as granules formulation	Enhances N accumulation, N <sub>2</sub> fixation, total biomass, nodule amount and weight	[67]
Burkholderia	Combination of substrates clay, rice bran and rock phosphate	High viability of beneficial bacteria.	[68]

### C. Pelletized Bio-fertilizer Formulation

Pellet formulation is one of the carrier-based bio-fertilizers. Pellet bio-fertilizer is a solid, compressed, and spherical particle of bio-fertilizer made from a compressed mixture of microorganisms and carrier matter. The principle of pellet formulation of bio-fertilizer is to apply force onto the biofertilizer formulation until it forms into pellets [70]. The basic formulation of pellet bio-fertilizer is almost similar to other formulations, starting with the selection and isolation of microbial strains of interest. In this formulation, the microbial inoculants require carrier material to provide solid-state and favorable conditions to the bio-fertilizer. The carrier material and inoculants are mixed, and the mixture is subsequently passed through a pellet press machine to create a small and pellet shape of bio-fertilizer.

Based on research made by [70], three sources of carrier substances which are aquatic weed, biochar, and compost, have been used to evaluate their efficacy in bio-fertilizer. The result showed that the application of pellet bio-fertilizer made from the combination of compost, bioinoculant, and nutrient supplement on paddy plants yielded the highest productivity in terms of shoot height, the number of grains per panicle, and weight of the grains. Also stated in [71], the pellet formulation of *Trichoderma harzianum* in the wheat carrier is effective as it can support the growth and productivity of mustard and tomato.

On the one hand, the formulation of pelletized bio-fertilizer using organic fertilizer substrate from chicken manure mixed with four bacterial strains has shown a significant effect on the host plant [72]. Moreover, as in [72], also proposed that the liquid form of bacteria culture is the most suitable for pellet bio-fertilizer compared to immobilized cells. Therefore, rather than mixing, the pelleted carrier may also be coated with microorganisms by spraying the liquid inoculant after the pelleting process. Figure 3 shows the carrier formulation to produce bio-fertilizer from rubberwood, decanter cake, rice husk ash, and spent coffee ground mixed with purple non-sulfur bacteria *Rhodopseudomonas palustris* and *Rubrivivax gelatinosus* [73]. Table IV summarised the pellet bio-fertilizer formulation.

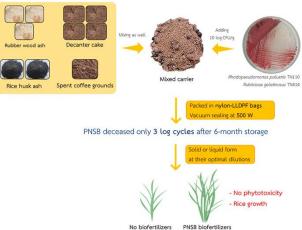


Figure 3. Pellet formulation of bio-fertilizer by using rubberwood, decanter cake, rice husk ash, and spent coffee ground mixed with purple non-sulfur bacteria *Rhodopseudomonas palustris and Rubrivivax gelatinosus* [73].

 TABLE IVV

 PELLET FORMULATIONS OF BIO-FERTILIZER

Bio-fertilizer/	Formulation	Result	Ref.
Strains			
Azospirillum sp., Pseudomonas fluorescens AMF	Different carrier substances • Biochar • Compost • Aquatic weed	Compost, bioinoculant, and nutrient supplement yielded the highest productivity.	[70]
Serratia marcescens (P2- 2& 8G), Enterobacter cloacae (P1-8), Klebsiella pneumoniae (CB36)	The bacteria are inoculated as liquid culture and incorporated into organic fertilizer substrates.	Pelleted liquid bio- fertilizer retains the bacteria population and improves the growth of the host.	[72]
Trichoderma harzianum	Wheat grains (pellet).	Benefit productivity of mustard and tomato.	[71]

#### D. Immobilized/Encapsulated Formulation of Bio-fertilizer

Immobilized bio-fertilizer is recognized as the advanced formulation of bio-fertilizer nowadays. It is a formulation where the microorganism cells are encapsulated and attached to inert and insoluble material. Encapsulation of bio-fertilizer was invented to boost up the cell's resistance against environmental stress and unfavorable soil condition [74]. As a result, it exhibits higher stability toward pH and temperature and is more resistant to environmental change [75]. Other than that, immobilization assists the gradual and consistent release of the microorganism or enzyme to the soil. Encapsulation may include macro and microform depending upon the intended application. However, the technology of encapsulation and microencapsulation in bio-fertilizer is still new and limited to the experimental in the laboratory. In this formulation, a prepared culture of bio-fertilizer is mixed with encapsulation matrix formulation (alginate) and formed into rigid capsules by ejecting dropwise of the mixture into calcium chloride. This immobilization technique may be employed to the microorganism itself or the enzyme produced from the fermentation by the microorganism (Table V). As [76] explained, alginate-based macrocapsules of bio-fertilizer offer better function and impact to the host plant compared to other formulations. In addition, the macrocapsules bio-fertilizer is preserved at a high number in field conditions and has a significantly longer shelf than the control without matrix.

Apart from that, there is also research performed regarding microencapsulation of bio-fertilizer. As [77] suggested, microencapsulation of rhizobacteria with gelatin polymer through spray drying methods is an appropriate formulation to enable efficient release of the bacteria. Furthermore, [75] stated that encapsulated bio-fertilizer could efficiently function in sodium alginate polymer with gelatin concentrations of 91.23% and 87.23% to protect potatoes from *Fusarium solani* disease.

Aside from bacteria, published research on immobilized enzymes for bio-fertilizer has the potential to be commercialized in agricultural practices. For example, according to [78], the immobilized keratinase enzyme can degrade chicken feathers into a bio-fertilizer to enhance the growth and chlorophyll content of *Solanum lycopersicum*.

TABLE V IMMOBILIZED/ENCAPSULATED FORMULATIONS OF BIO-FERTILIZER

Bio-fertilizer/ Strains	Formulation	Result	Ref.
Pseudomonas	Encapsulated in alginate	Optimum gelatin	[75]
fluorescens	beads with different gelatin concentrations.	concentration: 91.23% and 87.23%	
Azospirillum	<ul> <li>Macrocapsule</li> <li>Alginate (3%)</li> <li>Standard starch (44.6%)</li> <li>Modified starch (2.4%)</li> </ul>	The bacteria have longer shelf life.	[76]
Rhizobacteria	Microencapsulation within gelatin polymer.		
Immobilized keratinase enzyme	Entrapment in calcium- alginate beads. Chicken feather (organic material) is degraded by the enzyme.	s. improved the growth and	

Each formulation is able to deliver benefits to plant and crop productivity, but it may still have limitations (Table V1). Thus, the selection of suitable bio-fertilizer based on plant conditions will determine the successfulness of the inoculation. Appropriate methodology and detailed process are necessary to produce a good formulation of bio-fertilizer with competent strains of bacteria. Most of the studies have emphasized the advancement of bio-fertilizer formulations. In contrast, very limited work has been carried out on the optimization of biofertilizer quality and affectivity that could completely substitute chemical fertilizer in agriculture.

### **IV. CONCLUSIONS**

Microorganisms have an essential role in promoting plant growth. Plant probiotic bacteria are able to interact with a host by supplementing nutrients and compounds required by the plant. The introduction of bio-fertilizers has shown incredible effects on plants and is able to conserve nature. The selection of effective bacterial strains and suitable carriers is important in developing bio-fertilizers. The technology revolution today has contributed to the advanced development of bio-fertilizer that serves important benefits to the agricultural sector globally.

There are many studies that have been established to create improved formulations of bio-fertilizer. Therefore, this review focuses on an advanced bio-fertilizer formulation that can meet the objective of an effective bio-fertilizer. In the future, a study on the formulation of bio-fertilizer is needed to be carried out. These varieties of bio-fertilizer formulations could then be commercially practiced for more sustainable agriculture and a green environment.

TABLE VI ADVANTAGES AND DISADVANTAGES OF BIO-FERTILIZER FORMULATION

Types of	Bio-fertilizer	Advantage	Disadvantage
bio-	formulation		
fertilizer			
Inoculant	Liquid	Does not require	• Low
suspension	Liquiu	Does not require     carrier material	
suspension			stability during
		• Easy to inoculate.	application (no
		Suitable for soilless	protection from
		agriculture systems.	the carrier)
		<ul> <li>Microbial cells</li> </ul>	<ul> <li>Complicate</li> </ul>
		survive with the	d storage
		addition of the additive.	condition (at
		Contain a high	4°C)
		concentration of cells.	
		<ul> <li>Cost-effective</li> </ul>	
Carrier-	~ .	<ul> <li>Convenient for</li> </ul>	Selection of
based bio-	Granular	storage and application	compatible
fertilizer		<ul> <li>Easy to handle</li> </ul>	carrier material
	Pellet	(dust-free)	is difficult
		<ul> <li>Supply nutrients</li> </ul>	<ul> <li>Complex</li> </ul>
		for microorganisms	preparation of
		Provide a suitable	the carrier
		environment for	<ul> <li>Bulky in</li> </ul>
		microbes	size: High cost
		Carrier protect cells	for storage and
		from the chemical	transportation
		component	umsportation
		<ul> <li>Long shelf life</li> </ul>	
		Facilitate soil	Expensive
	Encapsulated	application	method
	/	Easy to store (room	Require
	Immobilized	-	*
	minoonized	<ul><li>temperature)</li><li>Ensure consistent</li></ul>	specific
			equipment
		release of	• Comulting
		microorganisms	Complicate
		depending on nutrient	d production
		availability in soil	• Not
		<ul> <li>Matrix: provide</li> </ul>	commercially
		reliable protection	available
		against unfavorable	(limited to
		conditions	laboratory)

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#### REFERENCES

- N. Rawat, S. Wungrampha, S.L. Singla-Pareek, M. Yu, S. Shabala & A. Pareek, (2022). "Rewilding staple crops for the lost halophytism: Toward sustainability and profitability of agricultural production systems," Molecular Plant, 15(1), 45-64. https://doi.org/10.1016/j.molp.2021.12.003
- [2] S. Komatsu, K. Saito & T. Sakurai, (2022). "Changes in production, yields, and the cropped area of lowland rice over the last 20 years and factors affecting their variations in Cote d'Ivoire," Field Crops Research 277: 108424. https://doi.org/10.1016/j.fcr.2021.108424
- [3] P. Temniranrat, K. Kiratiratanapruk, A. Kitvimonrat, W. Sinthupinyo & S. Patarapuwadol, (2021). "A system for automatic rice disease detection from rice paddy images serviced via a Chatbot," Computers and Electronics in Agriculture, 2021. 185: p. 106156. https://doi.org/10.1016/j.compag.2021.106156
- [4] Z. Jiang, Z. Dong, W. Jiang & Y. Yang, (2021). "Recognition of rice leaf diseases and wheat leaf diseases based on multi-task deep transfer learning," Computers and Electronics in Agriculture, 186: p. 106184. https://doi.org/10.1016/j.compag.2021.106184
- [5] A. Abbas, S. Jain, M. Gour & S. Vankudothu, (2021). "Tomato plant disease detection using transfer learning with C-GAN synthetic images," Computers and Electronics in Agriculture, 187: p. 106279. https://doi.org/10.1016/j.compag.2021.106279
- [6] P. Walerowski, A. Gundel, N. Yahaya, W. Truman, M.A. Sobczak, M. Olszak, S. Rolfe, L. Borisjuk & R. Malinowski, (2018). "Clubroot Disease Stimulates Early Steps of Phloem Differentiation and Recruits SWEET Sucrose Transporters within Developing Galls," The Plant Cell, 30(12): p. 3058-3073. https://doi.org/10.1105/tpc.18.00283
- [7] T.A.T. Pham, J.G. Schwerdt, N.J. Shirley, X. Xing, Y.S.Y. Hsieh, V. Srivastava & V. Bulone, (2019). "A. Little, Analysis of cell wall synthesis and metabolism during early germination of Blumeria graminis f. sp. hordei conidial cells induced in vitro," The Cell Surface, 5: p. 100030. https://doi.org/10.1016/j.tcsw.2019.100030
- [8] S. Castanheira, N. Mielnichuk & J. Perez-Martin, (2014). "Programmed cell cycle arrest is required for infection of corn plants by the fungus *Ustilago maydis*," Development, 141(24): p. 4817-4826. https://doi.org/10.1242/dev.113415
- [9] J. Han, Y. Dong & M. Zhang, (2021). "Chemical fertilizer reduction with organic fertilizer effectively improve soil fertility and microbial community from newly cultivated land in the Loess Plateau of China," Applied Soil Ecology, 165, 103966. https://doi.org/10.1016/j.apsoil.2021.103966
- [10] P. Vejan, T. Khadiran, R. Abdullah & N. Ahmad, (2021). "Controlled release fertilizer: A review on developments, applications and potential in agriculture," Journal of Controlled Release, 339, 321-334. https://doi.org/10.1016/j.jconrel.2021.10.003
- [11] S. Nadarajan, S. Sukumaran, F.B. Lewu, T. Volova, S. Thomas & K.R, Rakhimol (2021). Chapter 12 - Chemistry and toxicology behind chemical fertilizers. Controlled Release Fertilizers for Sustainable Agriculture pp. 195-229: Academic Press.
- [12] O.A. Fasusi, C. Cruz & O.O. Babalola, (2021). "Agricultural Sustainability: Microbial Biofertilizers in Rhizosphere Management," Agriculture, 11, 163. https://doi.org/10.3390/agriculture11020163
- [13] R. Ramachandran & J. Mourin, (2016). "Overview of the POPs pesticide situation in Malaysia," International POPs Elimination Project, pp. 1–15.
- [14] H. Rozita, (2011). "Chlorpyrifos blood level and exposure symptoms among paddy farmers in Sabak Bernam, Malaysia," International Journal of Public Health Research, vol.1, pp. 1–6, [Online]. Available at http://spaj.ukm.my/ijphr/index.php/ijphr/article/view/2.
- [15] N. Sharma & R. Singhvi, (2017). "Effects of Chemical Fertilizers and Pesticides on Human Health and Environment: A Review," International Journal of Agriculture, Environment and Biotechnology, vol. 10, pp. 675. doi: 10.5958/2230-732X.2017.00083.3

- [16] J. S. Singh, V. C. Pandey & D. P. Singh, "Efficient soil microorganisms: A new dimension for sustainable agriculture and environmental development," Agriculture, Ecosystems and Environment, vol. 140, pp. 339–353, 2011. https://doi.org/10.1016/j.agee.2011.01.017
- [17] E. Malusá & N. Vassilev, (2014) "A contribution to set a legal framework for biofertilizers," Applied Microbiology and Biotechnology, vol. 98, pp. 6599–6607. doi: 10.1007/s00253-014-5828v.
- [18] Y. Li, H. Li, X. Han, G. Han, J. Xi, Y. Liu, Y. Zhang, Q. Xue, Q. Guo & H. Lai, (2021). "Actinobacterial biofertilizer improves the yields of different plants and alters the assembly processes of rhizosphere microbial communities," Applied Soil Ecology, 2021. 171: p. 104345. https://doi.org/10.1016/j.apsoil.2021.104345
- [19] N. Zainuddin, M.F. Keni, S.A.S. Ibrahim, & M.M.M. Masri, (2021). "Effect of integrated biofertilizers with chemical fertilizers on the oil palm growth and soil microbial diversity," Biocatalysis and Agricultural Biotechnology, 39, 102237. https://doi.org/10.1016/j.bcab.2021.102237
- [20] M.S. Mahmud & K.P. Chong, (2021). "Formulation of biofertilizers from oil palm empty fruit bunches and plant growth-promoting microbes: A comprehensive and novel approach towards plant health," Journal of King Saud University - Science, 33(8), 101647. https://doi.org/10.1016/j.jksus.2021.101647
- [21] J. Sakpirom, T. Nunkaew, E. Khan & D. Kantachote, (2021). "Optimization of carriers and packaging for effective biofertilizers to enhance *Oryza sativa L.* growth in paddy soil," Rhizosphere, 19, 100383. https://doi.org/10.1016/j.rhisph.2021.100383
- [22] N. Bangash, S. Mahmood, S. Akhtar, M.T. Hayat, S. Gulzar & A. Khalid, (2021). "Formulation of biofertilizer for improving growth and yield of wheat in rain dependent farming system," Environmental Technology & Innovation, 24, 101806. https://doi.org/10.1016/j.eti.2021.101806
- [23] S. Tyagi, R. K. Naresh, S. Prakash, G. Yadav, S. Tiwari, B. Rawat & N. Sharma, (2019). "Conservation agriculture, bio-fertilizers and biopesticides: A holistic approach for agricultural sustainability and food security: A review," International Journal of Chemical Studies, vol. 7, pp. 3036–3046.
- [24] E. Menendez and P. Garcia-Fraile, (2017). "Plant probiotic bacteria: solutions to feed the world," AIMS Microbiology, vol. 3, pp. 502–524. doi: 10.3934/microbiol.2017.3.502
- [25] S. Kumar, Diksha, S.S. Sindhu & R. Kumar, (2021). "Biofertilizers: An ecofriendly technology for nutrient recycling and environmental sustainability," Current Research in Microbial Sciences, 3, 100094. https://doi.org/10.1016/j.crmicr.2021.100094
- [26] O.O. Babalola, (2010). "Beneficial bacteria of agricultural importance," Biotechnology Letters, 32(11), 1559-1570. https://doi.org/10.1007/s10529-010-0347-0
- [27] B. Essalimi, S. Esserti, L.A. Rifai, T. Koussa, K. Makroum, M. Belfaiza, S. Rifai, J.S.p. Venisse, L. Faize, N. Alburquerque, L. Burgos, S.E. Jadoumi & M. Faize, (2021). "Enhancement of plant growth, acclimatization, salt stress tolerance and verticillium wilt disease resistance using plant growth-promoting rhizobacteria (PGPR) associated with plum trees (*Prunus domestica*)," Scientia Horticulturae, 291, 110621. https://doi.org/10.1016/j.scienta.2021.110621
- [28] S. Rahimi, M. Talebi, B. Baninasab, M. Gholami, M. Zarei & H. Shariatmadari, (2020). "The role of plant growth-promoting rhizobacteria (PGPR) in improving iron acquisition by altering physiological and molecular responses in quince seedlings," Plant Physiology and Biochemistry, 155, 406-415. https://doi.org/10.1016/j.plaphy.2020.07.045
- [29] M. Boyer, R. Bally, S. Perrotto, C. Chaintreuil & F. Wisniewski-Dye, (2008). "A quorum-quenching approach to identify quorum-sensingregulated functions in *Azospirillum lipoferum*," Research in Microbiology, 159(9), 699-708. https://doi.org/10.1016/j.resmic.2008.08.003
- [30] S. Bukhat, A. Imran, S. Javaid, M. Shahid, A. Majeed & T. Naqqash, (2020). "Communication of plants with microbial world: Exploring the regulatory networks for PGPR mediated defense signalling," Microbiological Research, 238, 126486. https://doi.org/10.1016/j.micres.2020.126486
- [31] A. Hartmann, (2020). "Quorum sensing N-acyl-homoserine lactone signal molecules of plant beneficial Gram-negative rhizobacteria support plant growth and resistance to pathogens," Rhizosphere, 16, 100258. https://doi.org/10.1016/j.rhisph.2020.100258
- [32] I.-E. Marcano, C.-A. Diaz-Alcantara, B. Urbano, F. Gonzalez-Andres,(2016). "Assessment of bacterial populations associated with banana tree roots and development of successful plant probiotics for

banana crop," Soil Biology and Biochemistry, 99, 1-20. https://doi.org/10.1016/j.soilbio.2016.04.013

- [33] S. Kalam, A. Basu & A.R. Podile, (2020). "Functional and molecular characterization of plant growth promoting Bacillus isolates from tomato rhizosphere," Heliyon, 6(8), e04734. https://doi.org/10.1016/j.heliyon.2020.e04734
- [34] G. Qi, S. Chen, L. Ke, G. Ma & X. Zhao, (2020). "Cover crops restore declining soil properties and suppress bacterial wilt by regulating rhizosphere bacterial communities and improving soil nutrient contents," Microbiological Research, 238, 126505. https://doi.org/10.1016/j.micres.2020.126505
- [35] L. Xue, B. Sun, Y. Yang, B. Jin, G. Zhuang, Z. Bai & X. Zhuang, (2021). "Efficiency and mechanism of reducing ammonia volatilization in alkaline farmland soil using Bacillus amyloliquefaciens biofertilizer," Environmental Research, 202, 111672. https://doi.org/10.1016/j.envres.2021.111672
  [36] G. Santoyo, (2021). "How plants recruit their microbiome? New
- [36] G. Santoyo, (2021). "How plants recruit their microbiome? New insights into beneficial interactions," Journal of Advanced Research. https://doi.org/10.1016/j.jare.2021.11.020
- [37] N. Bahareh, N. BouaÃ-cha, J.S. Metcalf, S.J. Porzani & O. Konur, (2021). "Plant-cyanobacteria interactions: Beneficial and harmful effects of cyanobacterial bioactive compounds on soil-plant systems and subsequent risk to animal and human health," Phytochemistry, 192, 112959. doi: 10.1016/j.phytochem.2021.112959
- [38] P. Anubrata & D. Rajendra, (2014) "Isolation, characterization, production of bio-fertilizer and its effect on vegetable plants with and without carrier materials," International Journal of Current Research, vol. 6, pp. 7986–7995.
- [39] A. N. Yadav, P. Verma, B. Singh, V. S. Chauhan, A. Suman & A. K. Saxena, (2017) "Plant growth promoting bacteria: biodiversity and multifunctional attributes for sustainable agriculture," Advances in Biotechnology and Microbiology, vol. 5, pp. 1–16. doi: 10.19080/AIBM.2017.05.555671
- [40] T. Mahanty, S. Bhattacharjee, M. Goswami, P. Bhattacharyya, B. Das, A. Ghosh & P. Tribedi, (2017). "Bio-fertilizers: a potential approach for sustainable agriculture development," Environmental Science and Pollution Research, vol. 24, pp. 3315–3335.
- [41] S. M. Nadeem, Z. A. Zahir, M. Naveed, H. N. Asghar & M. Arshad, (2010). "Rhizobacteria capable of producing ACC-deaminase may mitigate salt stress in wheat," Soil Science Society of America Journal, vol. 74, pp. 533–542. https://doi.org/10.2136/sssaj2008.0240
- [42] R. Kumar, N. Kumawat & Y. K. Sahu, (2017). "Role of bio-fertilizers in agriculture," Pop Kheti, vol. 5, pp. 63–66.
- [43] J. U. Itelima, W. J. Bang, I. A. Onyimba, M. D. Sila & O. J. Egbere, (2018). "Bio-fertilizers as key player in enhancing soil fertility and crop productivity: A review," Direct Research Journal of Agriculture and Food Science, vol. 6, pp. 73–83.
- [44] C. Leaungvutiviroj, P. Ruangphisarn, P. Hansanimitkul, H. Shinkawa & K. Sasaki, (2010). "Development of a new bio-fertilizer with a high capacity for N<sub>2</sub> fixation, phosphate and potassium solubilization and auxin production," Bioscience, Biotechnology and Biochemistry, vol. 74, pp. 1098–1101. https://doi.org/10.1271/bbb.90898
- [45] M. Abdel-Salam & A. Shams, (2012). "Feldspar-K fertilization of potato (*Solanum tuberosum L.*) augmented by bio-fertilizer," Journal of Agriculture and Environmental Sciences, vol. 12, pp. 694–699. doi: 10.5829/idosi.aejaes.2012.12.06.1802
- [46] H. T. Tam & C. N. Diep, (2017). "Isolation and characterization of bacteria of mangrove rhizosphere in the Mekong Delta, Vietnam," International Journal of Innovations in Engineering and Technology, vol. 9.
- [47] J. Dighton, (2009) "Mycorrhizae," In Encyclopedia of Microbiology, Elsevier Inc. pp. 153–162. doi: 10.1016/B978-012373944-5.00327-8
- [48] P. I. Djighaly, N. Diagne, M. Ngom, D. Ngom, V. Hocher, D. Fall, D. Diouf, L. Laplaze, S. Svistoonoff & A. Champion, (2018). "Selection of arbuscular mycorrhizal fungal strains to improve *Casuarina equisetifolia L.* and *Casuarina glauca Sieb.* tolerance to salinity," Annals of Forest Science, vol. 75, pp. 1–11. doi: 10.1007/s13595-018-0747-1
- [49] E. Malusá, L. Sas-Paszt & J. Ciesielska, (2012). "Technologies for beneficial microorganisms inocula used as bio-fertilizers," The Scientific World Journal, 2012. https://doi.org/10.1100/2012/491206
- [50] Y. Bashan, (1998). "Inoculants of plant growth-promoting bacteria for use in agriculture," Biotechnology Advances, vol. 16, pp. 729–770. https://doi.org/10.1016/S0734-9750(98)00003-2
- [51] C. Badgley, J. Moghtader, E. Quintero, E. Zakem, M. J. Chappell, K. Aviles-Vazquez, A. Samulon & I. Perfecto, (2007). "Organic agriculture

and the global food supply," Renewable Agriculture and Food Systems, pp. 86–108. doi: 10.1017/S1742170507001640

- [52] D. Kantachote, K. Kowpong, W. Charernjiratrakul & A. Pengnoo, (2009). "Microbial succession in a fermenting of wild forest noni (Morinda coreia Ham) fruit plus molasses and its role in producing a liquid fertilizer," Electronic Journal of Biotechnology, vol. 12, pp. 9– 10. http://dx.doi.org/10.4067/S0717-34582009000300009
- [53] R. Anandham, N. Premalatha, H.J. Jee, H.Y. Weon, S.W. Kwon, R. Krishnamoorthy, P.I. Gandhi, Y.K. Kim & N.O. Gopal, (2015). "Cultivable bacterial diversity and early plant growth promotion by the traditional organic formulations prepared using organic waste materials," International Journal of Recycling of Organic Waste in Agriculture, 4(4), 279-289. https://doi.org/10.1007/s40093-015-0107-1
- [54] T. Phibunwatthanawong & N. Riddech, (2019). "Liquid organic fertilizer production for growing vegetables under hydroponic condition," International Journal of Recycling of Organic Waste in Agriculture, 8(4), 369-380. https://doi.org/10.1007/s40093-019-0257-7
- [55] E. L. D. Amalraj, G. P. Kumar, S. K. M. H. Ahmed, R. Abdul & N. Kishore, (2013). "Microbiological analysis of panchagavya, vermicompost, and FYM and their effect on plant growth promotion of pigeon pea (Cajanus cajan L.) in India." Organic Agriculture, vol. 3, pp. 23-29. https://doi.org/10.1007/s13165-013-0042-2
- [56] A.N. Nikitin, I.A. Cheshyk, G.Z. Gutseva, E.A. Tankevich, M. Shintani & S. Okumoto, (2018). "Impact of effective microorganisms on the transfer of radioactive cesium into lettuce and barley biomass," Journal of Environmental Radioactivity, 192, 491-497. https://doi.org/10.1016/j.jenvrad.2018.08.005
- [57] R. Anandham, N. Premalatha, H. J. Jee, H. Y. Weon, S. W. Kwon, R. Krishnamoorthy, P. I. Gandhi, Y. K. Kim & N. O. Gopal, (2015). "Cultivable bacterial diversity and early plant growth promotion by the traditional organic formulations prepared using organic waste materials," International Journal of Recycling of Organic Waste in Agriculture, vol. 4, pp. 279–289. https://doi.org/10.1007/s40093-015-0107-1
- [58] M. S. Santos, M. A. Nogueira & M. Hungria, (2019). "Microbial inoculants: reviewing the past, discussing the present and previewing an outstanding future for the use of beneficial bacteria in agriculture," AMB Express, vol. 9. https://doi.org/10.1186/s13568-019-0932-0
- [59] R. de Souza, A. Ambrosini & L. M. P. Passaglia, (2015). "Plant growthpromoting bacteria as inoculants in agricultural soils," Genetics and Molecular Biology, vol. 38, pp. 401–419. https://doi.org/10.1590/S1415-475738420150053
- [60] M. Rahman, M. Rahman, A. A. Sabir, J. A. Mukta, M. M. A. Khan, M. Mohi-Ud-Din, M. G. Miah, M. Rahman & M. T. Islam, (2018). "Plant probiotic bacteria Bacillus and Paraburkholderia improve growth, yield and content of antioxidants in strawberry fruit," Scientific Reports, vol. 8, pp. 1–11. https://doi.org/10.1038/s41598-018-20235-1
- [61] P. Chop, K. Hoe & K. A. Rahim, (2020). "Multifunctional liquid biofertilizer as an innovative agronomic input for modern agriculture," In Conference: Research and Development Seminar, pp. 1–4.
- [62] G. P. Santhosh, (2015). "Formulation and shelf life of liquid biofertilizer inoculants using cell protectants," International Journal of Researches in Biosciences, Agriculture and Technology, vol. 2, pp. 243–247.
- [63] P. Sahu & G. P. Brahmaprakash, (2016). "Formulations of Biofertilizers – Approaches and Advances," In D. P. Singh, H. B. Singh, and R. Prabha (Eds.), Microbial Inoculants in Sustainable Agricultural Productivity, New Delhi: Springer, pp. 179–198. https://doi.org/10.1007/978-81-322-2644-4\_12
- [64] S. Shanmugam, (2015). "Granulation techniques and technologies: recent progresses," BioImpacts: BI, 5(1), 55-63. doi: 10.15171/bi.2015.04
- [65] Y. Bashan, L. E. De-Bashan, S. R. Prabhu & J.-P. Hernandez, (2014). "Advances in plant growth-promoting bacterial inoculant technology: formulations and practical perspectives (1998–2013)," Plant and Soil, vol. 378, pp. 1–33. https://doi.org/10.1007/s11104-013-1956-x
- [66] L. Herrmann & D. Lesueur, (2013). "Challenges of formulation and quality of bio-fertilizers for successful inoculation," Applied Microbiology and Biotechnology, vol. 97, pp. 8859–8873. https://doi.org/10.1007/s00253-013-5228-8
- [67] G. W. Clayton, W. A. Rice, N. Z. Lupwayi, A. M. Johnston, G. P. Lafond, C. A. Grant & F. Walley, (2004). "Inoculant formulation and fertilizer nitrogen effects on field pea: Nodulation, N2 fixation and nitrogen partitioning," Canadian Journal of Plant Science, vol. 84, pp. 79–88. https://doi.org/10.4141/P02-089
- [68] R. Anandham, K. H. Choi, P. I. Gandhi, W. J. Yim, S. J. Park, K. A. Kim, M. Madhaiyan & T. M. Sa, (2007). "Evaluation of shelf life and

rock phosphate solubilization of Burkholderia sp. in nutrient-amended clay, rice bran and rock phosphate-based granular formulation," World Journal of Microbiology and Biotechnology, vol. 23, pp. 1121–1129. https://doi.org/10.1007/s11274-006-9342-y

- [69] V.F. Majaron, M.G. da Silva, R. Bortoletto-Santos, R. Klaic, A. Giroto, G.G.F. Guimaraes, W.L. Polito, C.S. Farinas & C. Ribeiro, (2020). "Synergy between castor oil polyurethane/starch polymer coating and local acidification by A. niger for increasing the efficiency of nitrogen fertilization using urea granules," Industrial Crops and Products, 154, 112717. https://doi.org/10.1016/j.indcrop.2020.112717
- [70] B. K. W. Pathirana & P. N. Yapa, (2020). "Evaluation of different carrier substances for the development of an effective pelleted biofertilizer for rice (*Oryza sativa L.*) using co-inoculated bacteria and arbuscular mycorrhizal fungi," Asian Journal of Biotechnology and Bioresource Technology, 1–10. doi: 10.9734/ajb2t/2020/v6i130070
- [71] M. M. Haque, G. Ilias & A. Molla, (2012). "Impact of Trichodermaenriched bio-fertilizer on the growth and yield of mustard (Brassica rapa L.) and tomato (Solanum lycopersicon Mill.)," The Agriculturists, vol. 10, pp. 109–119. https://doi.org/10.1007/s40003-012-0025-7
- [72] M. Stella, M. Theeba & Z. I. Illani, (2019). "Organic fertilizer amended with immobilized bacterial cells for extended shelf-life," Biocatalysis and Agricultural Biotechnology, vol. 20, pp.101248. https://doi.org/10.1016/j.bcab.2019.101248
- [73] J. Sakpirom, T. Nunkaew, E. Khan & D. Kantachote, (2021). "Optimization of carriers and packaging for effective biofertilizers to enhance *Oryza sativa L.* growth in paddy soil," Rhizosphere, 19, 100383. https://doi.org/10.1016/j.rhisph.2021.100383
- [74] N. Vassilev, M. Vassileva, A. Lopez, V. Martos, A. Reyes, I. Maksimovic, B. Eichler-Lobermann & E. Malusà, (2015). "Unexploited potential of some biotechnological techniques for bio-fertilizer production and formulation," Applied Microbiology and Biotechnology. Springer Verlag. https://doi.org/10.1007/s00253-015-6656-4
- [75] M. M. Pour, R. Saberi-Riseh, R. Mohammadinejad & A. Hosseini, (2019). "Investigating the formulation of alginate- gelatin encapsulated Pseudomonas fluorescens (VUPF5 and T17-4 strains) for controlling Fusarium solani on potato," International Journal of Biological Macromolecules, vol. 133, pp. 603–613. https://doi.org/10.1016/j.ijbiomac.2019.04.071
- [76] E. Ivanova, E. Teunou & D. Poncelet, (2005). "Alginate based macrocapsules as inoculants carriers for production of nitrogen biofertilizers. In Proceeding of the Balkan Scientific Conference of Biology. Plovdiv, Vol. 2005, pp. 90–108. https://doi.org/10.2298/CICEQ0601031I
- [77] N. Bharti, S. K. S. Sharma, S. Saini, A. Verma, Y. Nimonkar & O. Prakash, (2017). "Microbial plant probiotics: problems in application and formulation," In Probiotics and Plant Health, pp. 317–335, Springer. doi: 10.1007/978-981-10-3473-2\_13
- [78] A. C. Oluwaseun, P. Phazang & N. B. Sarin, (2018). "Production of ecofriendly bio-fertilizers produced from crude and immobilized enzymes from Bacillus subtilis CH008 and their effect on the growth of Solanum lycopersicum," Plant Archives, vol. 18, pp. 1455–1462.